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Effect of Biological Priming on Metabolomic and Molecular Changes in Response to Drought Stress in *Brassica juncea*

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Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

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ABSTRACT

Indian mustard, Brassica juncea is a leading oilseed crop in India and plays a crucial role in the agricultural as well as oilseed home marketing system of the country. High oil and phytosterols, glucosinolate content make it a more valuable crop, and it is used for edible oil production, poultry feed. There, however, is always one major problem during the growth of trees and their productivity; drought. This paper discusses the use of bio-priming to compound drought resistance in Indian mustard. Bio-priming, which entails the use of various microorganisms in seed treatment is therefore modern system of farming. In the process of germination and plant growth it enhances the seed germination rate, seedling vigor and overall plant health through various metabolomic and

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molecular mechanisms. Researchers have demonstrated that bio-priming containing elements such as Pseudomonas fluorescens, Bacillus subtilis, and Trichoderma harzianum faciliates the uptake of sulfur, growth and improvement of tolerance to abiotic stresses. Furthermore, by impacting on expression of stress associated genes, bio-priming enhances the activity of the WRKY transcription factor as well as the production of abscisic acid (ABA) and jasmonic acid (JA), leading to drought tolerance. These first results based on drought tolerance genes help to priorities future breeding programs dedicated to the improvement of drought resistant cultivars. Bio-priming also stimulated the antioxidant defense pathway and improved the ability of plants to cope with oxidation pressure arising from drought stress. Overall, bio-priming is a relatively cost-effective, environmentally considerate method for enhancing drought tolerance in Indian mustard, thus being beneficial for sustainable farming and food production.

Keywords: Abiotic stress; bio-priming; Brassica juncea; drought; metabolomics.

1. INTRODUCTION

Indian mustard, *Brassica juncea*, a member of the Brassicaceae family, is an important oilseed crop in India and plays a vital part in the country's agriculture. With an oil content ranging from 38% to 50% [1], it is highly valued and accounts for about 80% of the nation's total rapeseed-mustard production [2]. This important crop during the Rabi season is produced through interspecific hybridization and has a high oil content (37–42%) [1]. India is a major producer of rapeseed-mustard, which is the third most important oilseed crop in the world and makes a considerable contribution to the oilseed economy of the nation [3]. Mustard is well known for being high in bioactive substances that have antiinflammatory and antioxidant qualities. It also contains phytosterols, phenolic compounds, glucosinolates, and anti-carcinogenic substances [4]. India, which lies in third place behind China and Canada in the world's mustard production scenario, is an important participant. Over 6.23 million hectares of mustard are cultivated in India, producing 9.34 million tons at a productivity of 1499 kg/ha [5]. With 5.96 million hectares under cultivation and 8.32 million tonnes produced annually at a yield of 1397 kg/ha, the nation provides 28.3% of the world's rapeseed production [6]. In terms of the economy of oilseeds in India, mustard is grown in states such as Rajasthan, Uttar Pradesh, Madhya Pradesh, Punjab, Haryana, West Bengal, Assam, and Bihar [7]. It ranks second behind groundnut. Rajasthan is a prominent mustard-producing state that contributes significantly to India's overall mustard production, with Madhya Pradesh and Haryana following closely after [6]. Furthermore, while mustard is produced in other states in India, including Gujarat, Andhra Pradesh, Maharashtra, Karnataka, and Uttar Pradesh, Madhya Pradesh makes up a

substantial portion of the country's GDP [8]. Drought is one of the major obstacles to Indian mustard growth and development, affecting yield and productivity. India's agriculture is highly dependent on its climate, which leaves it open to water scarcity and decreased crop growth as a result of weak monsoons [9]. Drought-proofing techniques are therefore required because of its reduced ability to grow and produce seed in drought conditions [10]. The significance of drought tolerance indices in the selection of highyielding and drought-tolerant cultivars, such Pusa Jaikisan, which demonstrated resilience to drought stress while sustaining high yields, has been highlighted by research on a variety of mustard genotypes [11]. In rainfed regions that are vulnerable to drought, increasing mustard's tolerance to water stress through plant breeding and agronomic techniques is essential for longterm productivity [12]. Seed priming is a new generation technique that helps to resist the plants from abiotic stresses like drought.priming has various approaches, among which there is a method where we use living micro-organisms such as bacteria, fungi, called as bio-priming [13]. Biological priming improves a plant's resistance to a range of stresses by inducing particular metabolomic reactions and molecular pathways [14].

2. BIOLOGICAL PRIMING

Priming is a conventional technique that is typically employed for solid crop stands and synchronized seedling growth, but in recent years, it has become a powerful tool for sustainable agriculture [15]. There are different priming methods that effect the gemination pattern of mustard such as hydro-priming, osmopriming, hormonal priming and bio-priming [16]. Bio-priming, a seed treatment approach which combines a physiological seed hydration and biological seed inoculation with a beneficial organism in order to protect seed component of disease management is a novel seed treatment method [17]. By improving the adherence of bacteria to seeds, seed priming with living inoculums enhances rhizosphere colonization and increases plant resistance to unfavourable environmental conditions [18]. Biopriming, a practice where seeds are treated with biocontrol agents together with priming agents, has been used to control a number of soil- and seedborne diseases [17]. Chemical fungicides are not the only option for growers to control seed and seedling diseases; biological seed treatments offer an alternative option for this and hence ecofriendly and sustainable. Various bioinoculants such as *Pseudomonas fluorescens* and *Bacillus subtilis* are used as biopriming agents to increase the sulphur Use Efficiency in *Brassica juncea* [19]. Other than these bioagent *Trichoderma asperellum* Th-14 is also use for promoting the plant growth in Indian mustard [20]. Mycorrhizal fungus-assisted biopriming of mustard plants has demonstrated encouraging outcomes in terms of improving plant growth and disease resistance. Studies have shown that the use of mustard oil cake, *Pseudomonas fluorescens*, and mycorrhizal fungi like *Glomus mosseae* and *Scutellospora erythropa* can successfully treat root-rot disorders brought on by pathogens like *Macrophomina phaseolina* [21]. To increase the rate and uniformity of seed

emergence and reduce numerous seedborne diseases, seed priming has been employed either alone or in conjunction with suitable fungicides and/or biocontrol agent [22]. It has been demonstrated that biological priming, which involves soaking seeds in different substances such as gibberellic acid (GA), salicylic acid (SA), and abscisic acid (ABA), greatly increases the ability of plants such as rapeseed to withstand drought stress [23]. The processes involved in bio priming in plants help to improve seed germination, seedling vigour, and overall plant growth. When plant growth-promoting microorganisms (PGPMs) are used in the biopriming process, various mechanisms are triggered [24]. These include the production of siderophores, the solubilization or mobilisation of soil nutrients, the induction of plant growthpromoting activities, and the synthesis of defense-related enzymes, phytoalexins, and beneficial biochemicals [25]. Additionally, by promoting the synthesis of growth regulators, boosting nutrient uptake, and shielding plants from pathogens, bio priming with PGPMs like PGPR can result in increased seed quality, germination speed, vigour index, growth promotion, and disease resistance [26].

3. PROCEDURE FOR SEED BIOPRIMING

List of bio-priming agents used in mustard:

Table 1. Bio-priming agents used in mustard

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Fig. 1. Process of Biopriming of mustard seed

4. PHASES OF PRIMING AND EFFECTS

It is crucial for priming to involve a series of environmental stimuli [32]. The process is initiated with a triggering stimulus and goes up to the time when the plant is exposed to a challenging stress called priming phase [33]. During this phase, the plant is kept in a state of readiness for attack because the levels of main and secondary metabolites, phytohormones, Salicylic Acid (SA), and Jasmonic Acid (JA) fluctuate slightly [34]. Stress-hardened plants move to the second level of stress-hardening known as post-challenge primed state through a second stress which triggers defence response mechanisms promptly [35]. This includes de novo production of antimicrobial chemicals. A primed plant can revert to naïve state, however, if it inherits primed status from the parent plants, then plants can be trans generationally primed [36].

5. METABOLOMIC AND MOLECULAR CHANGES DUE TO BIO-PRIMING IN MUSTARD

Priming can be caused by natural or artificial substances such certain agrochemicals, or by

interactions between the host plants and helpful microbes such as rhizobacteria, mycorrhizal fungi, diseases, or virulent/avirulent microorganisms and plants that have these kinds of interactions experience long-lasting cellular and organismal reprogramming, and they molecularly "remember" these interactions [36]. Bio-priming in Indian mustard have metabolomic changes as it involves seed priming, research reveals that the use of bioinoculants such as *Bacillus subtilis* and *Pseudomonas fluorescens* promotes sulfur usage and its uptake with increased levels of S content and enzymes responsible for S mineralization [19]. Sulphur increases the amount of glucosinolate and the proportion of oil content, which is essential for the production of oil [37] A lack of S causes an imbalance in the intake of nutrients, which in turn causes the loss of chlorophyll, stunted growth, and decreased agricultural yields [38]. Hence *B. subtilis and P. fluorescens* promotes the sulphur level that increases the oil content in mustard. Moreover, seed born plant hormones like GA3 and SA play vital role in enhancing several biochemical attributes of mustard crops including chlorophyll process and oil percentage in seed [39]. Studies on growth promotion, disease suppression and biochemical changes in

response to bio control agents like *Trichoderma harzianum*, *Pseudomonas fluorescens*, *Bacillus subtilis* were shown to affect growth, disease and tissue biochemical composition like dry matter, phenols, sugars, lipids in the leaves of Indian mustard and alter protein quantity [40]. Treatment of tomato plants with Bacillus subtilis against Fusarium oxysporum has been shown to boost lipid content [41]. There is a demonstration after an experiment that in all cultivars of mustard, the total phenol concentration decreased after reaching a maximum at 60 DAS in healthy leaves and 80 DAS in ill leaves [42]. Furthermore, the studies on transgenic technology in *B. juncea* have also given fruitful results regarding the management of fatty acid environment in the seed, and also changing the nature of the seed itself to increase the yield of the oil [43]. The main aim of the present study involving two candidate genes was to reduce the VLCUFA (i. e. , C22: To this end, the levels of one of the undesirable fatty acids, namely, erucic acid, in the seed-oil of one Indian mustard cultivar *B. juncea* cv. PCR7, a high-yielding variety and resistant to lodging and podshattering were altered through genetic engineering to change the carbon flux in the metabolic pathway of fatty acid biosynthesis accompanied by the corresponding changes in the expression of the respective First, the effects of over-expression of the novel acyl-ACP thioesterase of FatB originated from the MbFatB gene from Diploknema (Madhuca) butyracea on the VLCUFA content were examined in order to shorten the acyl chain-elongation process and increase C16 and C18 FAs in the seed-oil [44]. This study concluded process to reduced erucic acid, increased beneficial fatty acids, and enhanced oil content. It has been noted that using plant growth-promoting bacteria in biopriming techniques is an eco-friendly way to increase seed germination in a variety of crops, highlighting the significance of microbial interactions in raising germination rates and seedling vigor [45]. The studies using RAPD markers established that various mustard lines and hybrid have distinct genetic patterns suggesting diversified genetic base and have prospect of utilization in breeding program [46]. In addition, polymorphic amplicons, alleles, and gene diversity among several Indian mustard genotypes have been discovered by genetic diversity assessments employing SSR markers, offering insights for the crop's genetic development [47]. The biochemical and molecular properties of Indian mustard, such as

the concentration of sinigrin, the isozyme patterns of peroxidase and esterase, the intensity of seed protein bands, and the genetic diversity determined by SSR and ISSR markers, can all be affected by biopriming [48]. Furthermore, the cloning and function identification of a promoter in Indian mustard has shown strong expression activity under various stress conditions, suggesting a potential role in gene regulation for stress responses [49]. When Indian mustard genotype (RH 781) was inoculated with *Albugo candida* led to a significant increase in ascorbic acid content, and the most significant changes in metabolite levels such as ascorbic acid and glutathione were observed in the 3rd leaf of both genotypes after 3 days of inoculation, with minimal changes in the 4th leaf compared to healthy leaves and other inoculated leaves on all days of inoculation [50]. Indian mustard's molecular alterations, which impact the genotypes' genetic variability and biochemical composition, are influenced by biopriming [51].

6. EFFECT AND RESPONSE TO DROUGHT DUE TO BIO-PRIMING ON INDIAN MUSTARD

Drought modifies a number of physiological and anatomical factors that are essential for mustard crop physiology to adapt to the drought. Drought stress causes mustard plants to lose fresh weight, dry weight, and abscisic acid content [52]. Drought stress also alters the protein composition, chlorophyll content, and activity of enzymes like catalase and peroxidase in mustard, resulting in physiological alterations that impact drought resistance [53]. Moreover, osmotic adaptations to display drought tolerance preserve the turgidity of cells in Brassica species, including mustard, emphasizing the significance of physiological features in maintaining resistance to water stress [54]. Biopriming with drought-tolerant isolates such as *Trichoderma harzianum*, improves osmoregulation and induces physiological protection against oxidative damage in Indian mustard [12]. *Trichoderma* as a seed treatment before sowing can induce tolerance to water stress in plants, highlighting its potential as a sustainable approach to improving crop resilience and could thrive in water stress conditions and positively impact mustard plant growth under such stress [55]. By delaying changes in net photosynthesis, stomatal conductance, and chlorophyll fluorescence brought on by drought, this strategy eventually

Fig. 2. Phases of priming

makes plants more resilient to water stress. It has also been discovered that using bioregulators such as glycinebetaine and salicylic acid can improve drought tolerance and growth in mustard crops by lessening the detrimental effects of water stress [56]. Moreover, it has been noted that induced drought circumstances cause mustard types to mount a defence response. This reaction results in elevated antioxidant, detoxifying enzyme, and biochemical parameter levels that improve resistance to drought stress and alternaria blight infection [57]. Furthermore, it has been researched that pretreating mustard seedlings with hydrogen peroxide activates defence mechanisms, increasing membrane stability and resistance to drought-induced oxidative damage [58]. It has been shown that biological priming—pre-treating plants like Indian mustard with hydrogen peroxide or β-aminobutyric acid (BABA) increases their resistance to drought. Research has indicated that the use of hydrogen peroxide priming can initiate defence mechanisms, initiate ROS and MG detoxification processes, and sustain essential enzyme functions and redox equilibrium, thereby equipping seedlings to endure drought-induced oxidative stress [59].
Similarly, BABA administration has been BABA administration has been associated with increased accumulation of ABA, which causes stomatal closure and stress gene expression to accelerate, improving plant resistance to drought stress without causing genetic changes [60]. Indian mustard genetic expression linked to drought resistance and

tolerance has been demonstrated to be positively impacted by bio priming, more especially by the use of bio stimulants such as Ascophyllum nodosum extract. It has been shown that the application of bio stimulants can suppress stressresponsive negative growth regulators, like RESPONSIVE TO DESICCATION 26 (RD26), while preserving the expression of markers of the cell cycle, such as HISTONE H4 (HIS4), to facilitate the active progression of the cell cycle under drought stress [23].

7. GENES RESPONSIBLE FOR DROUGHT TOLERANCE IN INDIAN MUSTARD

Various genes are that are either already present or incorporated in plant can result in drought tolerance.

The genes responsible for drought tolerance in Indian mustard (Brassica species) have been identified through various studies. Several unique genes involved in drought tolerance have been identified by comparative transcriptome analysis, including genes related to photosynthesis, biosynthesis of glutathione, biosynthesis of IAA signal transduction, biosynthesis of amino acids, cysteine, and methionine, and glucosinolate [11]. These genes are involved in the molecular aspects of drought stress in Brassica species and also useful in the selection of droughtresistant cultivars for the improvement of drought tolerance in breeding programs [65]. Further, genes associated with protein phosphatase 2C

SI. No.	Gene	Importance	Reference
	BjRD26	A NAC transcription factor gene (NAM, ATAF1/2, and CUC2) implicated in the regulation of genes associated to stress and the response to drought stress.	[61]
	BjHSP17.8	This gene helps prevent damage to cellular proteins by encoding a little heat shock protein that is produced during drought stress.	[62]
3.	BjDREB2A	A transcription factor gene known as dehydration- responsive element-binding (DREB) that controls the expression of genes sensitive to stress in drought-prone environments.	[63]
	BjERF1	A gene that responds to ethylene and is involved in the regulation of genes that are sensitive to drought.	[64]

Table 2. Different genes responsible for drought tolerance in mustard

have been reported to be involved in drought stress and hence, it becomes important to know the genetic makeup of drought tolerance in Indian mustard [66]. IgWRKY50 and IgWRKY32, the genes of Iris germanica, are two genes which were found to be important for increasing drought resistance in Indian mustard [67]. The genes belonging to this superfamily of WRKY transcription factor are regulated by kind of stress conditions including PEG-6000, high temperature and ABA and reaches at its maximum level at 3 h in case of PEG-6000 treated plants, when overexpressed in both Arabidopsis plants, enhanced shooting with improved osmotic tolerance, higher germination rates, increased average root length, and increased stomatal closure and reduced water loss rates under drought stress conditions [67]. Moreover, the expression of the stressresponsive genes in the transgenic plants was also upregulated, proline content and soluble protein content in the plants also increased, soluble protein content in the plants also increased, together with SOD, POD and CAT activities, all of which accumulated in the transgenic plant to enhance the drought resistance of the plants [68]. Therefore, IgWRKY50 and IgWRKY32 genes could be prospective candidates for effective utilization in molecular breeding programs directed at improving DT in Indian mustard. Transgenic mustard lines demonstrated increased antioxidant enzyme activity, decreased oxidative stress markers, and improved physiological traits, leading to better survival and performance under drought conditions. Over-expression of the chickpea Metallothionein1 (MT1) gene in mustard significantly enhances drought tolerance [69].

8. EFFECT OF BIO-PRIMING ON THE EXPRESSION OF GENES INVOLVED IN DROUGHT TOLERANCE IN INDIAN MUSTARD

Research have shown that bio-priming
upregulates stress-responsive genes and upregulates stress-responsive signalling pathways, including those involved in ABA and JA production, which in turn affects the expression of genes linked to drought tolerance [70]. Furthermore, it has been discovered that bio-priming preserves the shoot apical meristem's function and encourages active cell cycle progression during drought stress, which enhances plant resilience and growth [59]. Through the activation of stress memory systems, reinforcement of cellular defence responses, and induction of phytohormone synthesis, bio priming plays a critical role in improving drought tolerance in plants. Research has demonstrated that, in drought-stressed environments, seed biopriming with bacteria resistant to drought can greatly increase crop yields [71]. Numerous seed priming methods, such as bio-priming with bacteria that promote plant growth, have been found to be effective ways to increase crops, resistance to stress, which will boost growth, dry matter accumulation, and overall plant performance in both ideal and drought-prone environments [72]. It has been discovered that bio priming with particular bacterial strains raises antioxidant levels, including those of catalase, superoxide dismutase, and ascorbic peroxidase, all of which are essential in reducing the harmful consequences of oxidative stress brought on by dehydration [73]. Additionally, it may result in the activation of genes linked to plant hormone signalling, cell wall remodelling, and stress defence, all of which improve the plant's overall drought tolerance in Indian mustard and increase its capacity to fend off the impacts of drought [74]. CabHLH10, a bHLH transcription factor that regulates yield attributes under drought stress by altering the expression of drought-responsive genes and photosynthetic efficiency genes, is one of the specific genes bio-primed for better drought tolerance in Indian mustard [75]. The development of drought-tolerant cultivars of Brassica species has also been aided by the identification of traits such as primary branches plant-1, secondary branches plant-1, siliquae plant-1, seeds siliqua-1, seed yield plant-1, and 1000-seed weight as trustworthy selection criteria for drought tolerance [40]. Additionally, differentially expressed genes involved in photosynthesis, glutathione biosynthesis, IAA signal transduction, amino acid biosynthesis, and ABA signal transduction pathways were found in Brassica napus through comparative transcriptome analysis. These genes support drought tolerance mechanisms in rapeseed cultivars [76]. Higher seed vigour index and germination energy under drought stress processes have been observed in rapeseed cultivars that use drought-tolerant bacteria for biopriming, such as the strain SH-8, to boost drought tolerance and germination potential in wheat seeds [77]. The desirable genes could be enhanced by molecular markers like RAPD and SSRs; which can be used to investigate the genetic diversity among Indian mustard genotypes. This allows for the identification of diverse parents with high heritability and genetic advance, which can be used to improve yield and quality in early segregating generations [78]. Enhancing genes associated with plant growth and seed germination is one of the many advantages of bio priming. Based on research, biopriming entails introducing advantageous microbes, including Bacillus species, into seeds to enhance gene expression and seedling growth [79]. The processes of seed biopriming include the release of defence-related enzymes, induction of plant growth-promoting activities, and solubilization of soil nutrients, all of which support increased gene expression and plant growth [24]. Furthermore, bio priming is regarded as a sustainable method of mitigating abiotic stresses- drought in crops while guaranteeing uniform stand development under stress situations [80]. It is an environmentally benign substitute for chemical fungicides. Overall, bio priming improves seed germination and seedling establishment in difficult settings by upregulating

genes involved in antioxidant defence and DNA damage repair at the molecular level [81].

9. CONCLUSION

Brassica juncea, also known as Indian mustard, is an important oilseed crop that plays a major role in the agricultural economy and food security of India. The crop's economic and health benefits are further enhanced by its high oil content and significant bioactive components. However, the crop's development and productivity are negatively impacted by the drought, which presents serious issues. Biological priming has become a viable approach to lessen these difficulties, including bio-priming with advantageous bacteria. This method induces particular metabolomic and molecular changes that improve seed germination, seedling vigour, and overall plant growth. It has been demonstrated that bio-priming increases drought tolerance through upregulating genes that respond to stress, improving physiological and biochemical characteristics, and encouraging advantageous interactions between microbes and plants. Research indicates that bio-priming can boost Indian mustard's resistance to drought stress, maintaining productivity and enhancing crop quality. These efforts are further supported by genetic and molecular breakthroughs that increase the crop's resistance to harsh conditions, such as the discovery of genes resistant to drought and the application of transgenic technology. As a whole, there is a lot of potential to increase Indian mustard's productivity and resistance to drought by combining biological priming techniques with conventional breeding and genetic methods. This integrated approach guarantees this essential crop's sustained economic and nutritional contributions to India's agricultural sector in addition to promoting sustainable agriculture.

DISCLAIMER (ARTIFICIAL INTELLIGENCE)

I, Aishmita Gantait, hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc) and text-to-image generators have been used during writing or editing of manuscripts.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- 1. Singh VV, Garg P, Meena HS, Meena ML. Drought stress response of Indian mustard (*Brassica juncea L*) genotypes; 2018.
- 2. Rajput RK, Singh S, Varma J, Rajput P, Singh M, Nath S, Ravindra C, Rajput K. Effect of different levels of nitrogen and Sulphur on growth and yield of Indian mustard (*Brassica juncea (L.)* Czern and Coss.) in salt affected soil. \sim 1053 \sim Journal of Pharmacognosy and Phytochemistry. 2018;7(1).
- 3. Kumar S, Yadav KG, Patel S, Singh P, Kumar N. Nutrient management improves nutrition, quality and economic feasibility of Indian Mustard (*Brassica juncea L*.). International Journal of Current Microbiology and Applied Sciences. 2020;9 (7):735–746.

Available:https://doi.org/10.20546/ijcmas.2 020.907.085

- 4. Poyil, Theertha. Bio active compounds of mustard, its role in consumer health and in the development of potential functional foods. Current Nutrition and Food Science. 2023;19(9):950-960.
- 5. Bhardwaj S, Solanki NS, Nagar C. Study of phenological parameters and agrometeorological indices of Indian Mustard (*Brassica juncea L. Czern*) Varieties under Different Sowing Dates. International Journal of Current Microbiology and Applied Sciences. 2020;9 (11):3431–3436.

Available:https://doi.org/10.20546/ijcmas.2 020.911.409

6. Basavanneppa MA, Kumar BA. Production and economic efficiencies as influenced by mustard genotypes in paddy fallows of Tungabhadra command Area of Karnataka. International Journal of Current Microbiology and Applied Sciences. 2020; 9(6):2376–2380.

Available:https://doi.org/10.20546/ijcmas.2 020.906.291

- 7. Sahu A, Salam JL, Verma S, Samuel S. Combining ability and Heterosis for seed yield and its attributing traits in Indian Mustard (*Brassica juncea L*. Czern and amp; Coss). International Journal of Current Microbiology and Applied Sciences.2020;9(7):720–727. Available:https://doi.org/10.20546/ijcmas.2 020.907.083
- 8. Shukla P, Gupta JK. Study of cost and profit strategies for the mustard crop in

Madhya Pradesh. Current Journal of Applied Science and Technology. 2020;79– 87.

Available:https://doi.org/10.9734/cjast/2020 /v39i2730922

- 9. Nivetha N, Kiruthika A, Asha AD, Lavanya AK, Vikram KV, Manjunatha BS, Paul S. Osmotolerant Rhizobacteria improve seedling Vigour and plant growth of mustard under water scarcity. International Journal of Current Microbiology and Applied Sciences. 2020;9(12):1928–1937. Available:https://doi.org/10.20546/ijcmas.2 020.912.229
- 10. Singh VV, Beer B, Meena HS, Kulshrestha S, Dubey M, Gurjar N, Garg P, Meena ML, Rai PK. Heterosis and combining ability analysis for yield and yield attributes in Indian Mustard (*Brassica juncea L.*). International Journal of Microbiology and Applied Sciences. 2020;9 (3):1622–1632. Available:https://doi.org/10.20546/ijcmas.2

020.903.190

- 11. Kumari vedna. Assessment of drought tolerance using drought tolerance indices and their inter relationship in mustard; 2020.
- 12. Sahoo CR, Dash M, Acharya N. Biochemical responses of Indian mustard to water stress. International Journal of Current Microbiology and Applied Sciences.2019;8(02):1711–1718. Available:https://doi.org/10.20546/ijcmas.2 019.802.201
- 13. Kępczyński J, Kępczyńska E. Plantderived smoke and Karrikin 1 in seed priming and seed biotechnology. In Plants. MDPI. 2023;12(12). Available:https://doi.org/10.3390/plants121 22378

14. Irshad K, Shaheed Siddiqui Z, Chen J, Rao Y, Hamna Ansari H, Wajid D, Nida K, Wei X. Bio-priming with salt tolerant endophytes improved crop tolerance to salt stress via modulating photosystem II and antioxidant activities in a sub-optimal environment. Frontiers in Plant Science. 2023;14.

Available:https://doi.org/10.3389/fpls.2023. 1082480

15. Paul S, Dey S, Kundu R. Seed priming: An tool towards sustainable agriculture. Plant Growth Regul. 2022;97: 215–234. Available:https://doi.org/10.1007/s10725- 021-00761-1

16. Thapa S, Baral B, Shrestha M, Chandra Dahal Dr K. Effect of different priming methods on germination behaviour of broadleaf mustard cv. Marpha Chauda Paate. Tropical Agrobiodiversity. 2022;3(2): 52–59.

Available:https://doi.org/10.26480/trab.02.2 022.52.59

- 17. Reddy PP. Bio-priming of Seeds. In Recent advances in crop protection. Springer India. 2012;83–90. Available:https://doi.org/10.1007/978-81- 322-0723-8_6
- 18. Prasad SR, Kamble UR, Sripathy KV, Bhaskar KU, Singh DP. Seed bio-priming for biotic and abiotic stress management. In Microbial Inoculants in Sustainable Agricultural Productivity: Research Perspectives. Springer India. 2016a;1:211– 228.

Available:https://doi.org/10.1007/978-81- 322-2647-5_12

19. Singh S, Sarkar D, Rakesh S, Singh RK, Rakshit A. Biopriming with bacillus subtilis enhanced the Sulphur use efficiency of Indian mustard under graded levels of Sulphur fertilization. Agronomy. 2023;13 (4).

Available:https://doi.org/10.3390/agronomy 13040974

- 20. Rawat, Monika, Rawat L, BT, CS. Comparative studies of the effect of microbial inoculants and inorganic chemicals on growth, yield, 247 yield contributing traits and disease suppression in two varieties of mustard green (*Brassica juncea L*.) under open field conditions in mid hills of U., and Pantnagar Journal of Research. 2022;20(2).
- 21. Neeraj, Singh K. *Cyamopsis tetragonoloba L* Taub inoculated with arbuscular mycorrhiza and Pseudomonas fluorescens and treated with mustard oil cake overcome Macrophomina root-rot losses. Biology and Fertility of Soils. 2010;46(3): 237–245. Available:https://doi.org/10.1007/s00374-

009-0422-7 22. Monika, Sood, Vipul, Kumar, Ruby, Rawal.

Seed biopriming a novel method to control seed borne diseases of crops. Null. 2021;181-223. DOI: 10.1016/B978-0-12-822919-4.00008-

9

23. Holness S, Bechtold U, Mullineaux P, Serino G, Vittorioso P. Highlight induced transcriptional priming against a

subsequent drought stress in Arabidopsis thaliana. International Journal of Molecular Sciences. 2023;24(7).

Available:https://doi.org/10.3390/ijms24076 60₈

- 24. Deshmukh AJ, Jaiman RS, Bambharolia RP, Patil VA. Seed biopriming-a review. International Journal of Economic Plants. 2020;7(1):038-043.
- 25. Rhaman MS, Imran S, Rauf F, Khatun M, Baskin CC, Murata Y, Hasanuzzaman M. Plants Seed Priming with Phytohormones: An Effective Approach for the Mitigation of Abiotic Stress; 2020. Available:https://doi.org/10.3390/plants100 10
- 26. Mitra D, Pellegrini M, Olatunbosun AN, Mondal R, Del Gallo M, Chattaraj S, Panneerselvam P. Seed priming with microbial inoculants for enhanced crop yield. In Microbial Inoculants. Academic Press. 2023;99-123.
- 27. Jovičić-Petrović J, Karličić V, Petrović I, Ćirković S, Ristić-Djurović JL, Raičević V. Biomagnetic priming—possible strategy to revitalize old mustard seeds. Bioelectromagnetics. 2021;42(3):238–249. Available:https://doi.org/10.1002/bem.2232 8
- 28. Chauhan A, Ranjan A, Jindal T. Biological Control Agents for Sustainable Agriculture, Safe Water and Soil Health. 2018;71– 83. Available:https://doi.org/10.1007/978-3-

319-58415-7_6

- 29. Sree BK, Bara BM, Pal AK. Effect of various presowing seed treatments on growth and yield parameters of Yellow Mustard (*Sinapis alba. L*) Variety (ISP - 186). International Journal of Environment and Climate Change. 2022;837–843. Available:https://doi.org/10.9734/ijecc/2022 /v12i1030871
- 30. Gupta J. Efficacy of Antagonists on mycelium growth and carpogenic germination of sclerotia of *Sclerotinia sclerotiorum* on Indian mustard. International Journal of Agricultural Invention. 2016;1(02):190–193. Available:https://doi.org/10.46492/ijai/2016. 1.2.12
- 31. Nayak US, Mishra I, Mishra BK. Field evaluation of bio-intensive IPM modules against important insect pests of mustard under North Coastal Plain Zone of Odisha. In Journal of Biological Control. 2014; 28(4).

32. Tenenboim H, Brotman Y. Omic relief for the Biotically stressed: Metabolomics of plant biotic interactions. In Trends in Plant Science. Elsevier Ltd. 2016;21(9):781– 791.

Available:https://doi.org/10.1016/j.tplants.2 016.04.009

33. Hilker M, Schwachtje J, Baier M, Balazadeh S, Bäurle I, Geiselhardt S, Hincha DK, Kunze R, Mueller-Roeber B, Rillig MC, Rolff J, Romeis T, Schmülling T, Steppuhn A, Van Dongen J, Whitcomb SJ, Wurst S, Zuther E, Kopka J. Priming and memory of stress responses in organisms lacking a nervous system. Biological Reviews.2016;91(4):1118–1133.

Available:https://doi.org/10.1111/brv.12215

34. Balmer A, Pastor V, Gamir J, Flors V, Mauch-Mani B. The "prime ome": Towards a holistic approach to priming. In Trends in Plant Science. Elsevier Ltd. 2015;20(7): 443–452.

Available:https://doi.org/10.1016/j.tplants.2 015.04.002

35. Shi L, Tang X, Tang G. GUIDE-Seq to detect genome-wide double-stranded breaks in plants. In Trends in Plant Science. Elsevier Ltd. 2016;21(10):815– 818.

Available:https://doi.org/10.1016/j.tplants.2 016.08.005

- 36. Tugizimana F, Mhlongo MI, Piater LA, Dubery IA. Metabolomics in plant priming research: The way forward? In International Journal of Molecular Sciences. MDPI AG. 2018;19(6). Available:https://doi.org/10.3390/ijms19061 759
- 37. Saleem M, Elahi E, Gandahi AW, Bhatti SM, Ibrahim H, Ali M. Effect of Sulphur application on growth, oil content and yield of sunflower. Sarhad Journal of Agriculture. 2019;35(4). Available:https://doi.org/10.17582/JOURN

AL.SJA/2019/35.4.1198.1203

- 38. Waraich EA, Hussain A, Ahmad Z, Ahmad M, Barutçular C. Foliar application of sulfur improved growth, yield and physiological attributes of canola (*Brassica napus L*.) under heat stress conditions. Journal of Plant Nutrition. 2021;45(3):369–379. Available:https://doi.org/10.1080/01904167 .2021.1985138
- 39. Tulsi, Ram, Yadav, Pradip, Kumar, Saini, Ravi, Yadav, Raj, Bhahadur. Effect of seed priming with plant growth regulators on physiological changes of Indian mustard

(*Brassica juncea L*. Czern and Coss.). Journal of Pharmacognosy and Phytochemistry. 2018;7:355-358.

- 40. Sharma S, Singh J, Munshi GD, Munshi SK. Biochemical changes associated with application of biocontrol agents on Indian mustard leaves from plants infected with Alternaria blight. Archives of Phytopathology and Plant Protection. 2010;43(4):315–323. Available:https://doi.org/10.1080/03235400 701804109
- 41. Ghonim MI. Induction of systemic resistance against Fusarium wilt in tomato by seed treatment with the biocontrol agent Bacillus subtilis; 1999.
- 42. Gupta SK, Gupta PP, Yadava TP, Kaushik CD. Metabolic changes in mustard due to Alternaria leaf blight. Indian Phytopathology. 1990;43(1):64-69.
- 43. Sinha S, Jha JK, Maiti MK, Basu A, Mukhopadhyay UK, Sen SK. Metabolic engineering of fatty acid biosynthesis in Indian mustard (*Brassica juncea*) improves nutritional quality of seed oil. Plant Biotechnology Reports. 2007;1(4):185– 197.

Available:https://doi.org/10.1007/s11816- 007-0032-5

44. Jha JK, Maiti MK, Bhattacharjee A, Basu A, Sen PC, Sen SK. Cloning and functional expression of an acyl-ACP thioesterase FatB type from Diploknema (Madhuca) butyracea seeds in Escherichia coli. Plant Physiology and Biochemistry. 2006;44(11– 12):645–655.

Available:https://doi.org/10.1016/j.plaphy.2 006.09.017

- 45. Rahaman S, Ramana C, Rao AS, Reddy BR. Field evaluation and economic analysis of manual drawn rotor Weeder for small farms. Economic Affairs (New Delhi). 2022;67(4):415–421. Available:https://doi.org/10.46852/0424- 2513.4.2022.7
- 46. Gami RA, Chauhan RM, Parihar A, Solanki SD, Kanbi VH. Molecular characterization in mustard [*Brassica juncea L*. Czern and Coss.]. International Journal of Agriculture, Environment and Biotechnology. 2013;6: 61-67.
- 47. Shrivastav A, Tripathi MK, Tiwari S, Tripathi N, Tiwari PN, Bimal SS, Rajpoot P, Chauhan S. Evaluation of genetic diversity in Indian Mustard (*Brassica juncea var.* rugosa) Employing SSR Molecular Markers. Plant Cell Biotechnology and

Molecular Biology. 2023;10–21. Available:https://doi.org/10.56557/pcbmb/2 023/v24i3-48245

- 48. Chaudhary JN. Biochemical and molecular
characterization of Indian mustard characterization of Indian mustard [*Brassica juncea (L.)* Czern and Coss] Genotypes (Doctoral dissertation, AAU, Anand); 2014.
- 49. Lang, Minglin An, Chunju Wu, Dingding Chu, Yuan, Song He, Zeng, Liu. Constitutive high-expression strongpriming Indian mustard promoter BjCET1and application thereof. Null; 2017.
- 50. Sapna1 SJVJ, ASR. Biochemical changes in antioxidant pathway metabolites in Indian mustard leaves due to white rust infection; 2011.
- 51. Shakeel A, Khan AA, Ahmad G. The potential of thermal power plant fly ash to promote the growth of Indian mustard (*Brassica juncea*) in agricultural soils. SN Applied Sciences. 2019;1(4). Available:https://doi.org/10.1007/s42452- 019-0404-9
- 52. Fang S, Zhao P, Tan Z, Peng Y, Xu L, Jin Y, Wei F, Guo L, Yao X. Combining physio-biochemical characterization and transcriptome analysis reveal the responses to varying degrees of drought stress in *Brassica napus L*. International Journal of Molecular Sciences. 2022;23 (15).

Available:https://doi.org/10.3390/ijms23158 555

- 53. Zhu J, Cai D, Wang J, Cao J, Wen Y, He J, Zhao L, Wang D, Zhang S. Physiological and anatomical changes in two rapeseed (*Brassica napus L*.) genotypes under drought stress conditions. Oil Crop Science. 2021;6(2):97–104. Available:https://doi.org/10.1016/j.ocsci.20 21.04.003
- 54. Chandra K, Kumar A. Impact of Water Deficit Condition on Osmoregulation of the Brassica Species; 2022. Available: www.irj.iars.info
- 55. Sharma KK, Singh US. Induction of water stress tolerance of mustard plants using Trichoderma as biological seed treatment. In Journal of Applied and Natural Science. 2014;6(2).

Available:www.ansfoundation.org

56. Singh A, Meena RS. (n.d.). Effect of foliar spray of Bioregulators and irrigation on dry matter accumulation of mustard (*Brassica juncea L*.).

- 57. Mallick SA, et al. Augmentation of biochemical defense against Alternaria blight of mustard by induction of drought stress. Journal of Plant Pathology. JSTOR. 2017;99(1):47–60. Available:http://www.jstor.org/stable/44280 572. Accessed 16 June 2024.
- 58. Mohammad, Anwar, Hossain, Masayuki, Fujita. Hydrogen peroxide priming stimulates drought tolerance in Mustard (*Brassica juncea L*.) Seedlings. 2013;4. DOI: 10.5376/PGT.2013.04.0020
- 59. Rasul F, Gupta S, Olas JJ, Gechev T, Sujeeth N, Mueller-Roeber B. Priming with a seaweed extract strongly improves
drought tolerance in Arabidopsis. drought tolerance in Arabidopsis. International Journal of Molecular Sciences. 2021;22(3):1–28. Available:https://doi.org/10.3390/ijms22031 469
- 60. Fleming TR, Fleming CC, Levy CCB, Repiso C, Hennequart F, Nolasco JB, Liu F. Biostimulants enhance growth and drought tolerance in Arabidopsis thaliana and exhibit chemical priming action. Annals of Applied Biology. 2019;174(2):153–165. Available:https://doi.org/10.1111/aab.12482
- 61. Tripathi P, et al. Functional characterization of Brassica juncea BjSOS3 and BjSOS2 homologs in plant salt tolerance. Frontiers in Plant Science. 2017;650. DOI: 10.3389/fpls.2017.00650
- 62. Yadav SK, et al. Heat shock proteins in Brassica juncea: Genome-wide identification, phylogeny, structure, and expression profiling under heat stress. Journal of Experimental Botany. 2019;70 (18):4771-4786. DOI: 10.1093/jxb/erz216
- 63. Luo Y, Dong D, Su Y, Wang X, Peng Y, Peng J, Zhou C. Transcriptome analysis of Brassica juncea var. tumida Tsen responses to *Plasmodiophora brassicae* by the biocontrol strain Zhihengliuella aestuarii. Functional and Integrative Genomics. 2018;18(3):301– 314.

Available:https://doi.org/10.1007/s10142- 018-0593-0

- 64. Divya K, et al. Ectopic expression of an ABA- and stress-inducible protein from Brassica juncea confers dehydration and salt tolerance to transgenic tobacco. Plant Cell Reports. 2010;29(5):459-469. DOI: 10.1007/s00299-010-0835-3
- 65. Sharma A, Kumari V, Rana A. Genetic variability studies on drought tolerance

using agro-morphological and yield contributing traits in rapeseed-mustard. International Journal of Bio-resource and Stress Management. 2022;13(7):771-779.

66. Zhu B, Xu H, Guo X, Lu J, Liu X, Zhang T. Comparative analysis of drought responsive transcriptome in Brassica napus genotypes with contrasting drought tolerance under different potassium levels; 2022a.

Available:https://doi.org/10.21203/rs.3.rs-2077417/v1

- 67. Zhang J, Huang D, Zhao X, Zhang M, Wang Q, Hou X, Di D, Su B, Wang S, Sun P. Drought-responsive WRKY transcription factor genes IgWRKY50 and IgWRKY32 from Iris germanica enhance drought resistance in transgenic Arabidopsis. Frontiers in Plant Science. 2022;13. Available:https://doi.org/10.3389/fpls.2022. 983600
- 68. Kumari A, Avtar R, Jattan ANM, Rani B. Screening for drought tolerance in Indian mustard (*Brassica juncea L*.) genotypes based on yield contributing characters and physiological parameters. Journal of Oilseed Brassica. 2019;10(1):1-7.
- 69. Lal S, Kumar V, Gupta U, Sushma, Shirke PA, Sanyal I. Overexpression of the chickpea Metallothionein 1 (MT1) gene enhances drought tolerance in mustard (*Brassica juncea* L.). Plant Cell, Tissue and Organ Culture (PCTOC). 2024;157(1):6.
- 70. Wang X, Li Q, Xie J, Huang M, Cai J, Zhou Q, Dai T, Jiang D. Abscisic acid and jasmonic acid are involved in drought priming-induced tolerance to drought in wheat. Crop Journal. 2021;9(1):120–132. Available:https://doi.org/10.1016/j.cj.2020.0 6.002
- 71. Lastochkina O, Yakupova A, Avtushenko I, Lastochkin A, Yuldashev R. Effect of seed priming with Endophytic bacillus subtilis on some physio-biochemical parameters of two wheat varieties exposed to drought after selective herbicide application. Plants. 2023;12(8). Available:https://doi.org/10.3390/plants120 81724
- 72. Cig F, Erman M, Inal B, Bektas H, Sonkurt M, Mirzapour M, Ceritoglu M. Mitigation of drought stress in wheat by bio-priming by PGPB Containing ACC Deaminase Activity. Ataturk University Journal of Agricultural Faculty. 2022;53(1):51–57. Available:https://doi.org/10.54614/AUAF.20 22.972753

73. Kumar H, Verma P, John SA, Blaise D. (n.d.). Physiological, biochemical and molecular manifestations in response to seed priming with elicitors under drought in cotton.

Available:http://14.139.232.166/opstat/

74. Wang X, Chen J, Ge J, Huang M, Cai J, Zhou Q, Dai T, Mur LAJ, Jiang D. The different root apex zones contribute to drought priming induced tolerance to a reoccurring drought stress in wheat. Crop Journal. 2021;9(5):1088– 1097.

Available:https://doi.org/10.1016/j.cj.2020.1 1.008

75. Thakro V, Malik N, Basu U, Srivastava R, Narnoliya L, Daware A, Varshney N, Mohanty JK, Bajaj D, Dwivedi V, Tripathi S, Jha UC, Dixit GP, Singh AK, Tyagi AK, Upadhyaya HD, Parida SK. A superior gene allele involved in abscisic acid signaling enhances drought tolerance and yield in chickpea. Plant Physiology. 2023; 191(3):1884–1912. Available:https://doi.org/10.1093/plphys/kia

c550

76. Zhu B, Xu H, Guo X, Lu J, Liu X, Zhang T. Comparative analysis of drought responsive transcriptome in Brassica napus genotypes with contrasting drought tolerance under different potassium levels; 2022b.

Available:https://doi.org/10.21203/rs.3.rs-2077417/v1

77. Shaffique S, Imran M, Kang SM, Khan MA, Asaf S, Kim WC, Lee IJ. Seed Bio-priming of wheat with a novel bacterial strain to modulate drought stress in Daegu, South Korea. Frontiers in Plant Science. 2023; 14.

Available:https://doi.org/10.3389/fpls.2023. 1118941

78. Kumar B, Pandey A, Singh SK. Genetic diversity for agro-morphological and oil quality traits in Indian mustard (*brassica juncea L.* Czern and Coss). In N Save Nature to Survive. 2013;8(3).

Available:www.thebioscan.in

79. Rakshit A. Impact assessment of bio priming mediated nutrient use effciency for climate resilient agriculture. In Climate Change and Agriculture in India: Impact and Adaptation. Springer International Publishing. 2018;56–68. Available:https://doi.org/10.1007/978-3- 319-90086-5_6

Gantait et al.; J. Adv. Biol. Biotechnol., vol. 27, no. 8, pp. 1325-1338, 2024; Article no.JABB.121240

- 80. Prasad SR, Kamble UR, Sripathy KV, Bhaskar KU, Singh DP. Seed bio-priming for biotic and abiotic stress management. In Microbial Inoculants in
Sustainable Agricultural Productivity: **Agricultural** Research Perspectives. Springer India. 2016b;1:211–228. Available:https://doi.org/10.1007/978-81- 322-2647-5_12
- 81. Forti C, Shankar A, Singh A, Balestrazzi A, Prasad V, Macovei A. Hydropriming and biopriming improve *Medicago truncatula* seed germination and upregulate DNA repair and antioxidant genes. Genes. 2020;11(3). Available:https://doi.org/10.3390/genes110

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