

British Microbiology Research Journal 7(4): 180-192, 2015, Article no.BMRJ.2015.110 ISSN: 2231-0886



SCIENCEDOMAIN international www.sciencedomain.org

Evaluating the Effect of Gamma Irradiation and Steam Sterilization on the Survival and Growth of Composted Sawdust Fungi in Ghana

N. K. Kortei^{1*}, G. T. Odamtten^{1,2}, M. Obodai³, V. Appiah¹ and M. Wiafe-Kwagyan²

¹Department of Nuclear Agriculture and Radiation Processing, Graduate School of Nuclear and Allied Sciences, P.O.Box AE 1, Atomic, Accra, Ghana. ²Department of Botany, University of Ghana, P.O.Box 55, Legon, Ghana. ³Food Research Institute, Council for Scientific and Industrial Research, P.O.Box M 20, Accra, Ghana.

Authors' contributions

This work was carried out in collaboration between all authors. Authors NKK and GTO designed the study, performed the statistical analysis, wrote the protocol, and wrote the first draft of the manuscript and managed literature searches. Authors VA, MO and MWK managed the analyses of the study and literature searches. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/BMRJ/2015/16521 <u>Editor(s):</u> (1) Laleh Naraghi, Plant Disease Research Department, Iranian Research Institute of Plant Protection, Tehran, Iran. <u>Reviewers:</u> (1) Walid Fediala Abd El-Gleel Mosa, Plant Production Department, Faculty of Agriculture (Saba Basha), Alexandria University, Alexandria, Egypt. (2) Anonymous, Malaysia. Complete Peer review History: <u>http://www.sciencedomain.org/review-history.php?iid=991&id=8&aid=8567</u>

> Received 5th February 2015 Accepted 23rd February 2015 Published 24th March 2015

Original Research Article

ABSTRACT

The growth and survival of some fungi associated with composted and pretreated sawdust particularly for mushroom cultivation were investigated on two growth media; Cooke's and Oxytetracycline Glucose Yeast Extract (OGYE). Some fungi were isolated during the composting of sawdust over a period of 28 days as well as after pretreatment with gamma irradiation doses of 5, 10, 15, 20, 25 and 32 kGy and moist heat of $100\pm 2^{\circ}$ C for 2.5 hours. Fungal counts ranged 4.72-5.77 log₁₀ CFU/g and 3.4- 4.1 log₁₀ CFU/g respectively for both media. Both pretreatment methods effectively reduced (p<0.05) fungal counts by an average of 1.48 (irradiation) and 2.22 (steam) log-cycle reductions on OGYE while there was an average of 3.13 (irradiation) and 1.10 (steam)

log-cycle reduction on Cooke's. Corresponding radiation sensitivities (D_{10} values) of 5.94±2.06 kGy and 5.64±1.12 kGy were recorded for fungi on both media respectively. Five species belonging to three genera were isolated on OGYE and among the fungi were *Aspergillus niger*, *Aspergillus fumigatus*, *Aspergillus ustus*, *Mucor racemosus* and *Rhizopus stolonifer*. Ten species belonging to four genera were also isolated on Cooke's medium included *Aspergillus niger*, *Aspergillus fumigatus*, *Aspergillus flavus*, *Aspergillus ustus*, *Aspergillus ustus*, *Aspergillus terreus*, *Aspergillus niger*, *Aspergillus fumigatus*, *Aspergillus flavus*, *Aspergillus ustus*, *Aspergillus terreus*, *Aspergillus parasiticus*, *Aspergillus alutaceus*, *Rhizopus stolonifer*, *Fusarium oxysporum* and *Mucor racemosus*. There was an observed reduction (p<0.05) in species diversity after pretreatment. The presence of these microorganisms on sawdust also has serious repercussions on animal and human health.

Keywords: Composting; gamma radiation; sawdust; steam; D₁₀ value; fungi; mushroom.

1. INTRODUCTION

Sawdust which is a source of organic waste material is used extensively in horticulture, in soil amelioration, as a substrate for cultivation of mushrooms and in diverse applications. Nonetheless, sawdust attains its optimum utilization if it is well decomposed and the lignin content turned to humus. Improperly decomposed sawdust could result in nitrogen immobilization and produce some toxic substances which will affect plants, fungi, animals and humans [1].

Composting is a process of exothermic biological oxidation of various organic wastes in the presence of air involving microorganisms. Through microbial decomposition, the organic matter is stabilized, matured and deodorized into a product rich in humic substances that can be used as organic soil conditioner, easy to store and distribute [2,3]. Modern concept of environmental management relies on the recycling of wastes. In this perspective, composting appears to be a safe form of treatment of some wastes and the reclamation of the nutrients contained in them [4]. It is environmentally friendly because the process produces a marketable end-product that can be used as a soil conditioner and organic fertilizer. Composting is therefore regarded as a successful strategy for the sustainable recycling of organic wastes [5,6].

The active component mediating the biodegradation and conversion processes during composting is the resident microbial community, among which fungi play a vital role [7]. Therefore, optimization of compost quality is directly linked to the composition and succession of microbial communities in the composting process [8,9]. There is practically no substance existing in nature that is not used by one microorganism or

another [10]. It is therefore necessary to identify the microorganisms present in the different processes, as several different species of microbes are usually involved [11,12]. Microbes play key role in the process monitoring of microbial succession is important for the effective management of the composting process.

Since composting methods and different substrates are associated with different composition of a microbial community, monitoring of the resident microbial population in compost is essential to determine its quality and field of application [8,13].

Fungi use many carbon sources; mainly lignocellulosic polymers and can survive in extreme conditions. They mainly are responsible for compost maturation [14]. They also degrade complex polymers such as polyaromatic compounds or plastics and are being increasingly applied to bioremediate soils contaminated with a wide range of pollutants [15,16].

Steam pretreatment of sawdust for mushroom cultivation, is the most widely used method in Ghana [17,18]. It has become essential to explore other methods of pasteurization and improve efficiency of production. Gamma irradiations which have short wave length, high energy photons, and have deep penetrating power so could serve both as a decontaminating agent and a hydrolytic agent [19,20,21] for the bioconversion of lignocellulosic materials to produce mushrooms [21].

The objective of this paper is to investigate the fungal populations and species diversity associated with the composting process, gamma irradiated and steam / moist heat pretreated sawdust substrate principally for mushroom cultivation.

2. MATERIALS AND METHODS

2.1 Composting

The compost was prepared by the outdoor single phase solid waste fermentation. Fresh sawdust obtained from Timber market Accra was mixed and composted as described by [21]. The compost was then stacked into a heap of about 1.5 m high, 1.5 m long and 1.5 m wide. This heap was left for composting for 28 days with regular turning every 4 days. At weekly intervals samples, moisture content of the compost were adjusted to approximately 68 - 70% [22] and then supplemented with rice bran (12%) and lime (0.5%).

2.2 Irradiation and Pasteurization of Sawdust Compost Bags

A batch of bagged composted sawdust substrates stored in polypropylene packs of 25 x 18 cm dimension were sterilized with moist heat at a temperature of 98-100°C for 2.5 hours. Another batch stored in same packs were subjected to irradiation doses of 0 kGy, 5 kGy, 10 kGy, 15 kGy, 20 kGy, 25 kGy and 32 kGy at the Gamma Irradiation Facility of the Ghana Atomic Energy Commission using a Cobalt- 60 source (SLL-02, Hungary) at a dose rate of 1.7 kGy per hour in air . The absorbed dose was confirmed by ethanol-chlorobenzene (ECB) dosimetry. Each treatment was replicated 6 times.

2.3 Enumeration and Characterization of Mycoflora

The dilution plate technique was used in estimating fungal populations. About 10 g fresh weight of sample was placed in 250 ml Erlenmeyer flask containing 100 ml sterile distilled water. The mixture was shaken at 140 rev. /min in a Gallenkamp Orbital Shaker for 30 min. Aliquot (1ml) of the suspension was placed in sterile universal bottles (MaCartney tubes) containing 9 ml of 0.1% peptone, and was serially diluted up to 1:10³. The fungal population was enumerated on modified Cooke's medium [23] and OGYE incubated at 30-32°C for 3-5 days for species diversity. The fungi were identified using their microscopic, morphological and cultural characteristics as outlined by [24].

2.4 Determination of Occurrence Percentage of Fungi

The incidence of occurrence of the different fungal isolates was done by a modified formular

of [25,26]. The frequency of occurrence of the fungal pathogens from the sawdust was determined. The total number of each isolate in the sawdust sample was obtained against the total number of all the isolates in the sample screened.

The mean value of this gives the occurrence percentage as the following equation shows:

occurrence percentage =
$$X/N \times 100$$
 (1)

where X = total number of each isolate in the sample and N = total number of all the isolates in the sample.

2.5 D₁₀ Values Determination

The D_{10} value is the reciprocal of the slope of the exponential part of a survival curve. This value may also be obtained from equation (2). The data was subjected to regression analysis. The surviving fractions, \log_{10} (*N*/*N*₀) of microorganisms, was calculated and used as relative changes of their actual viable cell counts. The D_{10} values were calculated by plotting \log_{10} (*N*/*N*₀) against dose (*D*) according to the equation

$$D_{10} = \frac{\text{Radiation Dose (D)}}{\log_{10} (No-N)}$$
(2)

where *N*o is the initial viable count; *N* is the viable count after irradiation with dose *D*; *D* is the radiation dose [27,28,29]. The linear correlation coefficient (r^2) and the regression equations were also calculated.

2.6 Statistical Analysis

The values obtained for the fungal counts were done in the standard forms and then transformed to logarithmic values and subjected to analysis of variance (one way ANOVA) using SPSS (Illinois, USA) version 9 for Microsoft windows.

3. RESULTS AND DISCUSSION

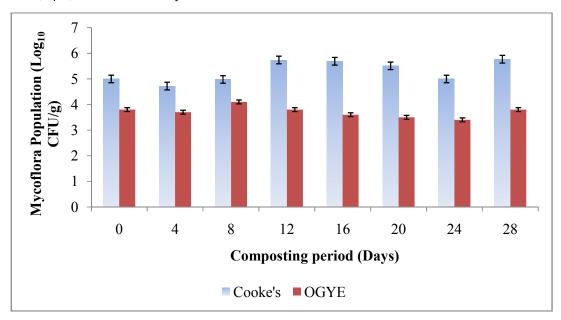
3.1 Fungal Population during Composting

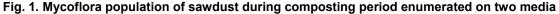
The dynamics of fungal community may be attributed generally to abiotic variables and nature of substrate [30]. There was variation of the fungal species in the sawdust with respect to the period of composting. The total mycoflora population enumerated on Cooke's medium was comparatively higher than on OGYE. The highest mycofloral population of 5.77 log₁₀ CFU/g was recorded for 28th day of composting while the lowest population of 4.72 log₁₀ CFU/g for the 4th day of composting. Statistically, mycoflora population of 0 (first), 4th, 8th and 24th days of composting which corresponded to 5.00, 4.72, 4.98 and 5.00 log₁₀ CFU/g respectively differed significantly (p<0.05) from 12th, 16th, 20th and 28th days of composting which corresponded to 5.74, 5.69, 5.51 and 5.77 log₁₀ CFU/g (Fig. 1). However, the highest mycoflora population enumerated on OGYE was 3.8 log₁₀ CFU/g for 0, 12th, 28thdays while the lowest recorded was 3.4 log₁₀ CFU/g for 24th day of composting. Composting days 0, 4, 12, 28 and 8corresponding to 3.8, 3.7, 3.8, 3.8 and 4.1 log₁₀ CFU/g respectively differed significantly (p<0.05) from days 16th and 20th corresponding to 3.6 and 3.5 \log_{10} CFU/g respectively which differed significantly (p<0.05) from 24th day of 4.1 \log_{10} CFU/g (Fig. 1).

Generally, at the beginning of composting, there was a comparatively low mycofloral population which suggested a minimal occurrence of factors such as moisture content, relative humidity and a wide fluctuation in air temperature which did not favour the sporulation of fungi on the sawdust substrate [31,32]. Microbial activities however increased as the composting period progressed. There was an increase in temperature, moisture content, pH, relative humidity and electrical conductivity of the activities according to [7] resulted in an increase in metabolic activities to produce enzymes by resident microorganisms to decompose the lignocellulosic materials. The enzymatic activities in compost piles are effective indicators for stress or adaptive practices of the microorganism different environmental to conditions, particularly to feed stock sources. Various hydrolytic enzymes can control the rate of decomposition of complex polymers during composting [33]. Obodai et al. [34] also suggested that the environmental and nutritional conditions created during composting might have selectively favored certain fungi to the detriment of others. This trend was in agreement with works of some researchers [34,35].

3.2 Effect of Gamma Irradiation and Steam Sterilization on Mycofloral Population of Sawdust Substrate

Both irradiation and steam techniques of pretreatment were effective in reducing the mycofloral population of sawdust. Although Comparatively high fungal counts were recorded on Cooke's medium than on OGYE medium (Fig. 2), gamma radiation dose of 5 kGy reduced the initial mycoflora of 5.77 \log_{10} CFU/g (control) by 0.68 log cycles to 4.89 \log_{10} CFU/g which did not differ (p>0.05) from the control. Doses 10 and 15 kGy recorded log cycle reductions of 1.97 and





2.50 respectively. Although their effectiveness did not differ (p>0.05), 5 and 10 kGy were similar. Doses 20, 25 and 32 kGy corresponded to log cycle reductions of 3.96, 4.68 and 4.76 respectively which showed no significant difference (p>0.05) in effectiveness but differed significantly (p<0.05) from the control, 5, 10 and 15 kGy.

Mycofloral population enumerated on OGYE followed a similar trend. Log cycle reductions of 0.5, 0.7, 1.0, 1.8, 2.2 and 2.7 were recorded for doses 5, 10, 15, 20, 25 and 32 kGy respectively (Fig. 2). Gamma radiation causes destruction and ultimately death to microorganisms by two mechanisms of action; first by direct action of producing free radicals by the ionizing energy and secondly by causing direct disruption of the DNA strand sequence which result in injury to the cell [36].

Steam/moist heat treated composted sawdust at temperature of 100±2°C was able to reduce mycofloral populations by 2.20 and 1.10 log cycles for Cooke's and OGYE respectively. The effectiveness of moist heat in reducing mycofloral populations were comparable (p>0.05) to gamma radiation doses of 5, 10 and 15 kGy on both media. However, doses 20, 25, and 32 kGy were more effective in mycofloral reduction by an average reduction range of 2.88, 3.44 and 3.73 log cycles. Reduction by these doses differed significantly (p>0.05) from moist heat effectiveness.

3.3 Occurrence Percentage of Microorganisms during Composting of Sawdust Substrate

Various mycoflora and microflora are involved in the composting of agricultural wastes and these range from fungi and bacteria that are thermophiles due to the temperature rise of 50-60°C, mesophiles and tolerant mesophiles.

Ten fungal species belonging to four genera namely: Aspergillus, Mucor, Rhizopus and encountered Fusarium were during the composting process. Generally, fungi increased in species diversity as composting proceeded in time (davs). On Cooke's medium, 0 day of composting recorded Aspergillus niger (38%), Aspergillus fumigatus (12%), Aspergillus ustus (15%), Aspergillus terreus (5%) and Rhizopus stolonifer (30%) (Fig. 3). There were a comparatively higher number of microbial counts and fungal species of observed on Cooke's than on OGYE.

On OGYE medium, five species belonging to three genera were enumerated. Fungi recorded were *A. niger* (30%), *A. ustus* (40%), *R. stolonifer* (30%) and *M. racemosus* (30%), *A. niger* (22.7%), *R. stolonifer* (47.3%) which corresponded to composting periods of 0 and 4 days respectively (Fig. 5).

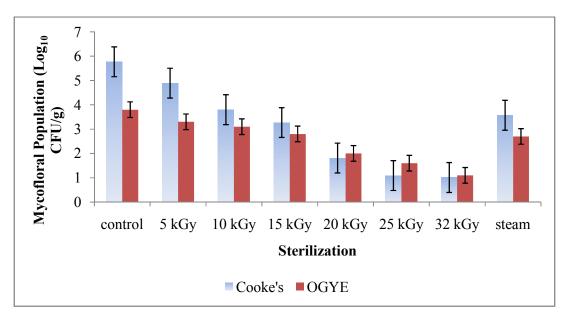


Fig. 2. Mycoflora population of sawdust after pretreatment cultured on two media

Results obtained imply a relatively high numbers of microorganisms were involved in the decomposition of the *T. scleroxylon* sawdust compost making it suitable for the growth of mushrooms as the diversity encourages different contributions biologically, chemically or physically [37,38].

The genus Aspergillus was predominant in all composting days of sawdust substrate. Several members of this genus have been reported by several researchers [39,40,41,42,43,44,45] to be capable of hydrolyzing the β -(1-4)-glucosidic linkage in the cellulose chain. According to [7], there is the production of metabolites in the compost during fungal growth which penetrates a cell and inhibits activity by chemical toxicity. Antagonism among fungi may be in the form of competition for nutrients, chemical antibiosis and lysis of mycelium. Antibiosis is the inhibition of one generation by the metabolic product of another. Although it is usually an inhibition of growth and sporulation, it may be toxic. Lysis is destruction and decomposition of biological materials by enzymes of the parasite. Fungal phenology observed in the compost may be partly attributed to antibiosis and lysis of mycelium.

Fungal species occurrence was irregular probably because the environmental and

nutritional conditions created during composting selectively favored certain fungi to the detriment of others.

Results obtained agreed with similar works reported by some researchers [34,46] as they investigated the various fungi associated with the composting of sawdust for mushroom cultivation.

3.4 Occurrence Percentage after Irradiation and Steam Sterilization

On Cooke's medium, the non-sterilized (control) sawdust substrate harbored *A. niger* (13.5%), *A. fumigatus* (26.5%), *A. flavus* (15%), *A. ustus* (10%), *A. parasiticus* (5%), *R. stolonifer* (25%), *M. racemosus* (5%). Conversely, radiation dose 5 kGy resulted in the retention of *A. niger* (18.5%), *A. fumigatus* (15.8%), *A. terreus* (10.7%), *A. ochraceus* (20%), *R. stolonifer* (10%) and *M. racemosus* (10.7%). While 10 kGy retained *A. niger* (20%), *A. fumigatus* (12%), *A. flavus* (15.7%), *A. ustus* (23.8%) and *A. parasiticus* (1.5%) (Fig. 4).

Dose 15 kGy, retained only species *A. niger* (24.5%), *A. fumigatus* (60%) and *R. stolonifer* (15.5%). Essentially, doses 20, 25 and 32 kGy retained no mycofloral species.

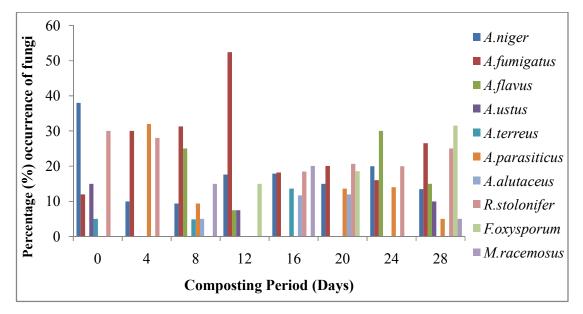


Fig. 3. Occurrence percentage of fungi of sawdust during composting period (Cooke's medium)

Kortei et al.; BMRJ, 7(4): 180-192, 2015; Article no.BMRJ.2015.110

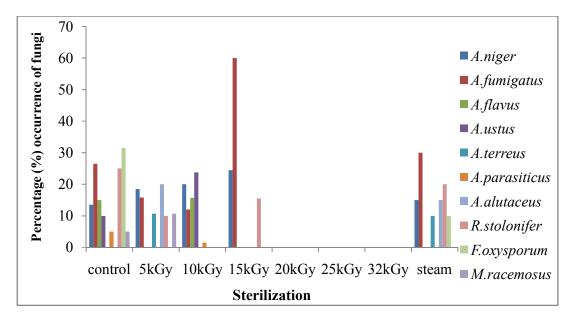
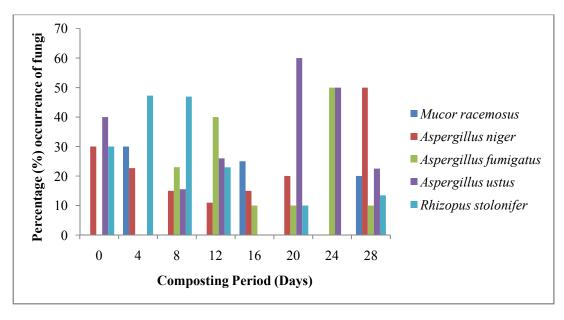
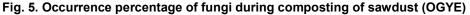


Fig. 4. Occurrence percentage of fungi after sterilization of sawdust (Cooke's medium)





Moist heat/steam sterilized sawdust retained *A. niger* (15%), *A. fumigatus* (30%), *A. terreus* (10%), *A. ochraceus* (15%), *R. stolonifer* (20%) and *F. oxysporum* (10%) (Fig. 4).

On OGYE medium, enumerated fungal species were comparatively minimal. The non- sterilized (control) retained *M. racemosus* (20%), *A. niger* (34.5%), *A. fumigatus* (27%), *A. ustus* (5%) and *R. stolonifer* (13.5%). Radiation doses 5 retained *M. racemosus* (18.2%), *A. niger* (30%), *A.*

fumigatus (50%) and *A. ustus* (1.8%). However 10 kGy retained only *A. niger* (100%) (Fig. 6).

Doses 15, 20, 25 and 32 kGy retained no fungi. However, steam retained *A. niger* (20%) and *A. fumigatus* (80%). The critical radiation target is the chromosomal DNA, damage to which inactivates the microorganism. About 90% of the damage is caused by OH- radicals released from the hydration layer around the DNA molecule. Usually the purine and pyrimidine bases are chemically changed and the phosphodiester backbone is broken (single-strand breaks) but some 5-10% double-strand breaks also occur [47]. Aquino [48] reported that radiation-resistant organisms are capable of withstanding effects on the plasma membrane of the cells. The rate of dose absorption has no effect on survival except when oxygen replenishment is involved. Elevated temperatures above 45°C are synergistically bactericidal because the higher temperatures damage the repair systems. To achieve this effect in spores, temperatures of 80 - 98°C are needed. Subfreezing temperatures raise the radiation resistance of vegetative cells as the water activity decreases and the diffusion of radicals is restricted.

3.5 Radiation Sensitivity (D₁₀ Values) of Fungi on Sawdust

sensitivity (the killing effect of Radiation in microorganisms is generally radiation) expressed by the decimal reduction dose or D₁₀ value [27] which was calculated from equation (1). The relative sensitivity of different microorganisms to ionizing radiation is based on their respective D₁₀ values (which is the dose required to reduce the population by 90%). Lower D₁₀ values indicate greater sensitivity of the organism in question. The data in Table 1. shows that gamma radiation doses achieved significant reduction in mycoflora during pasteurization of sawdust for mushroom production. On OGYE there was a good correlation coefficient of 0.971 with а corresponding mean D₁₀ value of 5.94±1.18 kGy (Fig. 7). Also on Cooke's medium, a good

correlation coefficient of 0.924 was established with a corresponding mean D_{10} value of 5.64±1.12 kGy was obtained (Fig. 7). This implies that a slightly higher mean dose of 5.94 kGy was required to inactivate fungi of composted sawdust. Hossain et al. [49] obtained a less D_{10} value of 0.43-1 kGy for yeasts and moulds. However, results were within range of values reported by Addo [50] for D_{10} values of fungi on irradiated flour (Hausa koko). Results obtained fell within the range of lethal ionization doses of 1.3-11 kGy and 4-11 kGy for moulds and yeasts respectively as reported by [51].

3.6 pH

The pH of fermenting substrate, initially recorded 7.39 which was neutral (Table 2). However, pH fluctuated to basic / alkaline (7-9) and then to acidic (6.81-8.04) as composting time increased. The drop in pH could probably be due to rapid breakdown of soluble and easily degradable carbon sources, resulting in a pH drop due to organic acids formation [52]. According to Tchobanoglous et al. [53], initial pH values of 7-7.5 is recommended as it aids production of lactic acid and acetic acids during initial degradation of biomass. Interestingly, at the 20th day of composting the pH value 9.17 was recorded which is in agreement with some researchers [54,55] who stated that in the thermophilic stage of composting, the pH could rise to 9 resulting in the release of ammonia, and there after the pH returns to near neutral conditions as the compost become mature. After pretreatment, pH ranged 7.45-8.39 (Table 3).

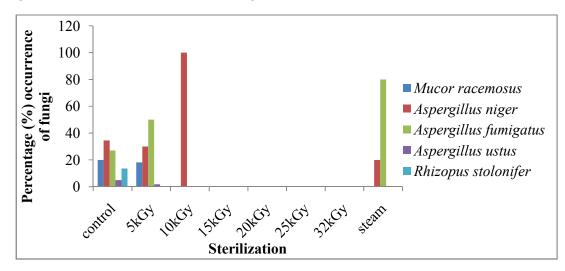


Fig. 6. Occurrence percentage of fungi after sterilization of sawdust (OGYE)

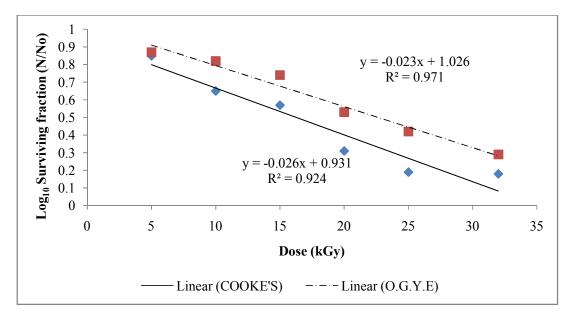


Fig. 7. Radiation sensitivity curves for fungi enumerated on OGYE and Cooke's media

Table 1. Mean D ₁₀ values of fungi associated with sawdust isolated from the two (2) growth	
media	

	Regression equation	r	D₁₀ value (kGy)
ke's	-0.026x + 0.931	0.924	5.64 ± 1.12
YE	-0.023x + 1.026	0.971	5.94 ± 1.18
		ke's -0.026x + 0.931 /E -0.023x + 1.026	ke's -0.026x + 0.931 0.924

 D_{10} values are means of 6 replicates \pm S.E

Table 2. Moisture content and pH readings of sawdust recorded during the composting period

Composting	Moisture content	рН
time (days)	(%)	
0	65.1 ^b	7.39 ^{ab}
4	60.3 ^a	8.04 ^b
8	58.2 ^a	6.81 ^a
12	61.1 ^a	8.20 ^b
16	65.8 ^b	8.75 ^b
20	64.9 ^b	9.17 ^c
24	59.7 ^a	8.93 ^b
28	62.8 ^a	8.85 ^b

Means with same superscripts in a column are not significantly different (P>0.05)

3.7 Moisture Content

Moisture content during the composting period ranged between 58.2 - 65.8%. Moisture is important in composting processes for two reasons: it facilitates substrate decomposition through mobilizing microorganism activities and also provides better conditions for nitrogen fixation in the compost. A low moisture content below a critical level (<30%), would decrease activities of microorganisms by restricting the motility and make them dormant [56]. Under drier conditions, the ammonium and ammonia present generate a higher vapor pressure; thus conditions are more favorable for nitrogen loss. On the other hand, a moisture content which is too high (>65%) could cause oxygen depletion and losses of nutrients through leaching [57].

Table 3. Effect of pretreatments on the pH and moisture contents of composted sawdust (*T. sceroxylon*)

Pretreatment	Moisture content (%)	рΗ
Steam	62.3 ^a	7.45 ^a
0 kGy	62.7 ^a	8.33 ^b
5 kGy	63.6 ^a	8.13 ^b
10 kĠy	64.1 ^a	8.29 ^b
15 kGy	63.7 ^a	8.39 ^b
20 kGy	64.0 ^a	8.08 ^b
24 kGy	63.0 ^a	8.36 ^b
32 kGy	61.8 ^ª	8.33 ^b

Means with same superscripts in a column are not significantly different (P>0.05)

4. CONCLUSION

From the results obtained, it was observed that fungal genera *Aspergillus, Mucor, Fusarium and Rhizopus* were involved in the decomposition of sawdust. The present study supports the idea that knowledge regarding species composition of the microorganisms of different composts can help to optimize compost quality.

Irradiation pretreatment with doses 20, 25 and 32 kGy were more effective in pasteurizing than steam. Doses 10 and 15 kGy were comparable to steam. Steam was however more effective than 5 kGy.

Researchers stand to benefit more from this research by acquiring more knowledge on the consortium of fungi that would facilitate the composting process to achieve the desired quality. Knowledge on pretreatment would enhance precision of elimination to reduce competition.

ACKNOWLEDGEMENTS

The authors are grateful to Mr. G. Akwetey of Mycology Unit, Department of Botany, University of Ghana. To Messers A. Azinkawe, M. Mensah and G. Agbeley of Food Research Institute, Council for Scientific and Industrial Research. Also to Messers S.N.Y Annan, E.A Quarcoo and S.W.N.O Mills of Radiation Technology Centre, Ghana Atomic Energy Commission, for their technical assistance.

COMPETING INTEREST

Authors have declared that no competing interests exist.

REFERENCES

- Rohanie B, Awg-Ahmad Sallehin AH. Feasibility of fungus bioaugmentation in sawdust composting. Proceedings of the 11th Symposium for the Malaysian Society of Applied Biology. Biological Balance Towards Life Sustainability. 2010;7-8.
- Domeizel M, Khalil A, Prudent P. UV spectroscopy: A tool for monitoring humification and for proposing an index of the maturity of compost. Bioresource Technology. 2004;94:177-184.
- 3. Gautam SP, Bundela PS, Pandey AK, Awasthi MK, Sarsaiya G. Composting of

municipal solid waste of Jabapur city, Global J. Environmental Res. 2010;4:43-46.

- Iranzo M, Canizares JV, Roca-Perez L, Sainz-Pardo I, Mormeneo S, Boluda R. Characteristic of rice straw and sewage sludge as composting materials in Valencia (Spain). Bioresource Technology. 2004;95(1):107-112.
- Fermor TR. Applied aspects of composting and bioconversion of lignocellulosic materials - An overview. International Biodeterioration & Biodegradation. 1993; 31:87–106.
- Tuomela M, Vikman M, Hatakka A, Itavaara M. Biodegradation of lignin in a compost environment: A review. Bioresource Technology. 2000;72:169– 183.
- Obodai M, Odamtten GT. Fungal phenology and attendant changes in agricultural lignocelluloses waste for mushroom cultivation: Status prospects and applications in food security. 2013;5-6.
- Peters S, Koschinsky S, Schwieger F, Tebbe CC. Succession of microbial communities during hot composting as detected by PCR-single-strandconformation polymorphism-based genetic profiles of small-subunit rRNA genes. Appl. Environ. Microbiol. 2000;66(03):930-936.
- Taiwo LB, Oso BA. Influence of composting techniques on microbial succession, temperature and pH in a composting municipal solid waste. African Journal of Biotechnology. 2004;3(4):239-243.
- Iranzo M, Sainz-Pardo I, Boluda R, Sánchez J, Mormeneo S. The use of microorganisms in environmental remediation. Ann. Microbiol. 2001;51:135-143.
- Hugenholtz P, Goebel BM, Pace NP. Impact of culture-independent studies on the emerging phylogenetic view of bacterial diversity. J. Bacteriol. 1998;180: 4765-4774.
- 12. Radajewski S, Inerson P, Parekh H, Murrell JC. Stable - isotope probing as a tool in microbial ecology. Nature. 2000; 403:646-649.
- Ishii K, Fukui M, Takii S. Microbial succession during a composting process as evaluated by denaturing gradient gel

electrophoresis analysis. J. Appl. Microbiol. 2000;89:768-777.

- Miller FC. Composting of municipal solid waste and its components. In: Microbiology of solid waste. (Eds.): AC Palmisano MA. Barlaz CRS Press. 1996;115-154.
- Minussi RC, de Moraws SG, Pastore GM, Durän N. Biodecolorization screening of microbial ecology. Nature. 2001;403:646-649.
- Ashraf R, Ali TA. Effect of oil (crude petroleum) on the survival and growth of soil. Appl. Microbiol. 2006;33:21-25.
- 17. Obodai M, Cleland- Okine J, Vowotor K.A. Comparative study on the growth and yield of *Pleurotus ostreatus* mushroom on different lignocellulosic by-products. J. Ind. Microbiol Biotechnol. 2003;30:146-149.
- Apetorgbor MM, Apetorgbor AK, Nutakor E. Utilization and cultivation of edible mushrooms for rural livelihood in Southern Ghana. 17th Commonwealth Forestry Conference, Colombo, Sri Lanka. 2005;1-20.
- 19. Gbedemah C, Obodai M, Sawyer LC. Preliminary investigations into the bioconversion of gamma irradiated agricultural wastes by *Pleurotus* spp. Radiation Phys. Chem. 1998;52(6):379-382.
- Mami Y, Peyvast G, Ziaie F, Ghasemnezhad M, Salmanpour V. Improvement of shelf-life and postharvest quality of white button mushroom by ⁶⁰CO γ-ray irradiation. Plant Knowledge Journal. 2013;2(1):1-7.
- Kortei NK, Odamtten GT, Obodai M, Appiah V, Annan SNY, Acquah SA, Armah JO. Comparative effect of gamma irradiated and steam sterilized composted 'wawa' (*Triplochiton scleroxylon*) sawdust on the growth and yield of *Pleurotus ostreatus* (Jacq.Ex.Fr.) Kummer. Innovative Romanian Food Biotechnology. 2014a;14:69-78.
- 22. Buswell JA. Potentials of spent mushroom substrates for bioremediation purposes. Compost, 1984;2:31-35.
- 23. Cooke WB. The use of antibiotics in media for the isolation of fungi from polluted water. Antibiotic and Chemotherapy. 1954; 4:657-662.
- 24. Samson AR, Hoekstra ES, Frisvad JC. Introduction of Food-Borne Fungi. 4th ed.

Netherlands: Pensen and Loogen. 1995; 12-20.

- Sampo S, Begero R, Buffa G, Lumpimosa, AM. Soil Fungi. Academic Press, London. 1997;6-27.
- 26. Ajiboye AE, Ajuwon IB, Adedayo MR. Physicochemical profile and microflora associated with the spoilage of soursop fruits (*Annona muricata*). Advances in Biotechnology Research. 2014;1(1):1-9.
- 27. Mohan A, Pohlman FW, Hunt MC. Inactivation of *E. coli* cells at low dose rates of gamma radiation. Arkansas Animal Science Dept. Report. 2011;120-123.
- Adu-Gyamfi A, Appiah V, Torgby-Tetteh W. Microbiological quality of chicken sold in Accra and determination of D₁₀-value of *E.coli*. Food and Nutrition Sciences. 2012; 3:693-698.
- 29. Kortei NK, Odamtten GT, Appiah V, Obodai M, Adu-Gyamfi A, Annan TA, Akonnor PT, Annan SNY, Acquah SA, Armah JO, Mills SWO. Microbiology quality assessment of gamma irradiated fresh and dried mushrooms (*Pleurotus ostreatus*) and determination of D10 values of *Bacillus cereus* in storage packs. European Journal of Biotechnology and Biosciences. 2014b;2(1):28-34.
- Thorman MN, Currah RS, Bayley SE. Succession of microfungal assemblages in the decomposing peat land plants. Plant Soil. 2003;250(3):323–333.
- Mc Tiernan KB, Couteaure MM, Berg B, Berg MP, de Anta RC, Kratz AGW, Pirussi P, Remacle J, Amalia V. Changes in chemical composition of *Pinus sylvestris* needle decomposition along a European coniferous forest climate transect. Soil Biol. Biochem. 2003;35(6):801–812.
- Cruz AG, Garcia SS, Rojas FJC, Ceballos AIO. Foliage decomposition of velvet bean during seasonal drought. Interciencia. 2002;27(11):625–630.
- Umsakul K, Dissara Y, Srimuang N. Chemical, physical and microbiological changes during composting of the water hyacinth. Pakistan Journal of Biological Science. 2010;13:985-992.
- 34. Obodai M, Amoa- Awua W, Odamtten, GT. Physical, chemical and fungal phenology associated with composting of 'wawa' sawdust (*Triplochiton scleroxylon*) used in the cultivation of oyster mushrooms in

Ghana. International Food Research Journal. 2010;17:229-237.

- Hubbe MA, Nahzad M, Sanchez C. Composting of cellulosics. Bioresources 2010;5(4):2808-2854.
- Abdel-Kadar A. Potential applications of ionizing radiations in postharvest handling of fresh fruits and vegetables. Food Technology. 1986;40(6):117-121.
- Tripothi JP, Yadar JS. Optimization of solid substrate fermentation of wheat straw into animal feed by *Pleurotus ostreatus*- A pilot effort. Animal Feed Science and Technology. 1992;37:59-72.
- Chiu SW, Law SC, Ching ML, Cheung KW, Chen MJ. Themes for mushroom exploitation in the 21st century: Sustainability, Waste management and Conservation. The Journal of General and Applied Microbiology. 2001;46:269-282.
- Steward C, Walsh JH. Cellulolytic activity of pure and mixed cultures of fungi. Trans.Br. Mycol. Soc. 1972;58:527.
- Thakur MP, Ram RN, Shukla CS. Mycoflora associated with paddy straw substrate during different stages and cropping months of oyster mushroom (*Pleurotus florida*). J. Mycol. Pl. Pathol. 2001;31(1):59-63.
- Ryckeboer J, Margaert J, Coosemans J, Deprins K, Swings J. Microbiological aspects of biowaste during composting in a monitored compost bin. Journal of Applied Microbiology. 2003;94:127-137.
- Iranzo M, J.V. Canizares, L. Roca-Perez, I. Sainz-Pardo, S. Mormeneo and R. Boluda. Characteristic of rice straw and sewage sludge as composting materials in Valencia (Spain). Bioresource Technology, 2004;95(1):107-112.
- Wouters IM, Spaan S, Douwes J, Doekes G, Heederik D. Overview of personal occupational exposure levels to inhale dust, endotoxin, β (1-3)-glucan and fungal extracellular polysaccharides in the waste management chain. Ann. Occup. Hyg. 2005;47:1-15.
- 44. Anastasi A, Varese GC, Marchisio VF. Isolation and identification of fungal communities in compost and vermicompost. Mycologia. 2005;97(1):33-44.
- 45. Pandey V, Sinha A. Mycoflora associated with decomposition of rice stubble mixed

with soil. Journal of Plant Protection Research. 2008;48(2):247-253.

- Omokaro O, Ogechi AA. Cultivation of mushroom (*Pleurotus ostreatus*) and the microorganisms associated with the substrate used. E- Journal of Science and Technology. 2013;8(4):49-59.
- Moseley B. Ionizing radiation action and repair. In: Mechanisms of action of food preservation procedures. Gould GW (Ed.). Elsevier Appl Sci, London. 1989;43-70.
- 48. Aquino S. Gamma radiation against toxigenic fungi in food, medicinal and aromatic herbs. In: Science against microbial pathogen: Communicating current research and technological advances. Mendelez-Vilas, A. (Ed.). 2011; 272-281.
- 49. Hossain F, Follet P, Dang Vu K, Salmieri S, Senoussi C, Lacroix M. Radiosensitization of *Aspergillus niger* and *Penicillium chrysogenum* using basil essential oil and ionizing radiation for food decontamination. Food Control. 2014;45: 156-162.
- 50. Addo AA. Mycological profile and aflatoxigenic potential of resident *Aspergillus* species of six packaged Ghanaian dehydrated foods. M. Phil. thesis submitted to the Department of Botany, University of Ghana; 2008.
- Frazier WC, Westhoff DC. Microbiología de los alimentos. 4th Zaragoza: Acribia. fungi. Int. J. Biol. Biotech. 1993;3(1):127-133.
- Beffa T, Blanc M, Lyon PF, Vogt G, Marchiana M, Fischer JL, Aragno M. Isolation of thermus strains from hot composts (60–80°C). Applied and Environmental Microbiology. 1996:62: 1723–1727.
- Tchobanoglous G, Theisen H, Vigil SA. Integrated solid waste management: engineering principles and management issues. 2nd Edn., McGraw-Hill International, New York, USA. 1993;11-15.
- 54. Hultman, J. Microbial diversity in the municipal composting process and development of detection methods, PhD thesis, Department of Ecological and Environmental Sciences, Faculty of Biosciences and Institute of Biotechnology and Vikki Graduate school in biosciences, University of Helsinki, Finland; 2009.

Kortei et al.; BMRJ, 7(4): 180-192, 2015; Article no.BMRJ.2015.110

- Ryckeboer J, Mergaert J, Coosemans J, Deprins K, Swings J. Microbiological aspects of biowastes during composting in a monitored compost bin. Journal of Applied Microbiology. 2003;94:127-137.
- 56. Hubbe MA, Nahzad M, Sanchez C. Composting of cellulosics. Bioresources. 2010;5(4):2808-2854.
- 57. Tiquia SM, Tam NFY, Hodgkiss I J. Microbial activities during composting of spent pig-manure sawdust litter at different moisture contents. Bioresource Technology. 1996;55:201-206.

© 2015 Kortei et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: http://www.sciencedomain.org/review-history.php?iid=991&id=8&aid=8567